Engineering particle trajectories in microfluidic flows using speckle light fields

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ABSTRACT

Optical tweezers have been widely used in physics, chemistry and biology to manipulate and trap microscopic and nanoscopic objects. Current optical trapping techniques rely on carefully engineered setups to manipulate nanoscopic and microscopic objects at the focus of a laser beam. Since the quality of the trapping is strongly dependent on the focus quality, these systems have to be very carefully aligned and optimized, thus limiting their practical applicability in complex environments. One major challenge for current optical manipulation techniques is the light scattering occurring in optically complex media, such as biological tissues, turbid liquids and rough surfaces, which give rise to apparently random light fields known as speckles. Here, we discuss an experimental implementation to perform optical manipulation based on speckles. In particular, we show how to take advantage of the statistical properties of speckle patterns in order to realize a setup based on a multimode optical fiber to perform basic optical manipulation tasks such as trapping, guiding and sorting. We anticipate that the simplicity of these “speckle optical tweezers” will greatly broaden the perspectives of optical manipulation for real-life applications.

Keywords: optical forces, Brownian motion, speckles, optical tweezers, light scattering, optical manipulation, random light fields

1. INTRODUCTION

Since their introduction in the 1970s \cite{1,2}, optical tweezers have been widely applied to non-invasively manipulate micro- and nano-objects, such as cells, organelles and macromolecules \cite{3-5}. They have, therefore, gained increasing importance as tools in microbiology and biophysics both for fundamental studies \cite{6} and for more advanced applications such as optical sorting and optical delivery \cite{3,7-8}. In particular, the development of techniques based on reconfigurable spatially extended patterns of light, such as multiple traps \cite{3,9-12} or periodic potentials \cite{13-17}, offers the promise of high throughput optical methods to be applied both in static and moving fluids. Also, particles’ delivery, trapping and manipulation over extended areas was demonstrated near a surface employing the evanescent fields associated, for example, to surface plasmons \cite{18} or to optical waveguides \cite{19}.

Most of current optical manipulation techniques rely either on carefully engineered optical systems or advanced fabrication tools. Although such conditions can be routinely met in research laboratories, similar requirements, sometimes very stringent, limit the applicability of these techniques, e.g., to biomedical and microfluidic applications, where simplicity, low-cost and high-throughput are paramount. Moreover, one major challenge common to all these techniques is the light scattering occurring in optically complex media, such as biological tissues, turbid liquids and rough surfaces, which naturally gives rise to apparently random light fields known as speckles \cite{20}. Earlier experimental works showed trapping of atoms and particles in a gas by high-intensity speckle light fields \cite{21-24}, while both static and time-varying speckle fields were related to the emergence of anomalous diffusion in colloids \cite{25-29}. Recently, we derived a theory to describe the motion of a Brownian particle in a speckle light field which allowed us to demonstrate numerically how a speckle field can be used to control the motion a Brownian particle in the limit of particles much smaller than the light wavelength (dipole approximation) \cite{29}. However, apart from these previous studies, the intrinsic

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randomness of speckle patterns is largely considered a nuisance to be minimized for most purposes in optical manipulation [30-31]. In fact, similar and even more complex effects have been extensively studied using periodic potentials rather than random potentials: these studies include the demonstration of guiding and sorting particles using either moving periodic potentials [16-17], static periodic potentials in microfluidic flows [7,11-12], or optical ratchets based on spatially symmetric energy landscapes [32-33]. Only recently we were able to demonstrate experimentally a novel technique for the collective manipulation of micrometer-sized particles in microfluidic flows based on extended static and time-varying speckle light field [34].

Here, we discuss an experimental implementation to perform optical manipulation based on speckles. In particular, we show how to take advantage of the statistical properties of speckle patterns in order to realize a setup based on a multimode optical fiber to perform basic optical manipulation tasks such as trapping, guiding and sorting. We anticipate that the simplicity of these “speckle optical tweezers” will greatly broaden the perspectives of optical manipulation for real-life applications.

2. EXPERIMENTAL IMPLEMENTATION

The speckle optical tweezers setup is schematically depicted in Figure 1(a). Aqueous dispersions of colloidal spheres are driven by a syringe pump with adjustable infusion flow rate (Harvard Apparatus Pump 11 Elite) through a microfluidic channel. The speckle light pattern for their optical manipulation (Figure 1(b)) is generated by coupling a laser beam (Coherent Verdi, maximum power 5W, $\lambda = 532$ nm) into a multimode optical fiber (core diameter 105 µm, NA = 0.22). The random appearance of speckle light patterns is the result of the interference of a large number of optical waves with random phases, corresponding to different eigenmodes of the fiber. More generally, speckle patterns can be generated by different processes: scattering of a laser on a rough surface, multiple scattering in an optically complex medium, or, like in this work, mode-mixing in a multimode fiber. The method chosen in this work provides some practical advantages over other methods, namely the generation of homogeneous speckle fields over controllable areas, flexibility and portability in the implementation of the device, as well as higher transmission efficiency. In our setup, the fiber output is brought in close proximity of the upper wall of the microfluidic channel by a micrometric two-axis mechanical stage that also guarantees the possibility of translating the speckle in the direction perpendicular to the fluid flow. Optical scattering forces push the particles in the direction of light propagation towards the lower wall of the microfluidic channel, so that they effectively confine the particles in a quasi two-dimensional space [1]. The particles are then tracked by digital video microscopy [35] on a color CMOS camera. The incoherent illumination for the tracking is provided by a LED at $\lambda = 625$ nm coupled into the same fiber using a dichroic mirror. Figure 1(c) shows the normalized spatial autocorrelation function of a typical speckle pattern interacting with the particles (Figure 1(b)), whose full width half maximum (FWHM = 2.20 ± 0.24 µm) provides an estimation of the average speckle grain size (Figure 1(d)), as defined by the diffraction process that generates the speckle pattern itself [20].
normalized spatial autocorrelation function, which (d) permits us to characterize the average speckle grain size as the FHWM of the autocorrelation along the axes (solid lines).

3. CONCLUSIONS

In conclusion, we have shown an experimental setup based on the use of a multimode optical fiber to perform optical manipulation of microparticles in microfluidic flows based on static, time- or space-varying speckle fields as in Reference [34]. Our technique, beyond demonstrating that random potentials are a valid alternative to more regular potentials for the purpose of optical manipulation, offers some additional advantages to current optical manipulation techniques, such as intrinsic robustness to noise, and to optical aberrations from the optical systems and the sample. The experimental results obtained with this technique can be understood and interpreted in terms of numerical simulation based on Brownian dynamics [29,36-37].

REFERENCES